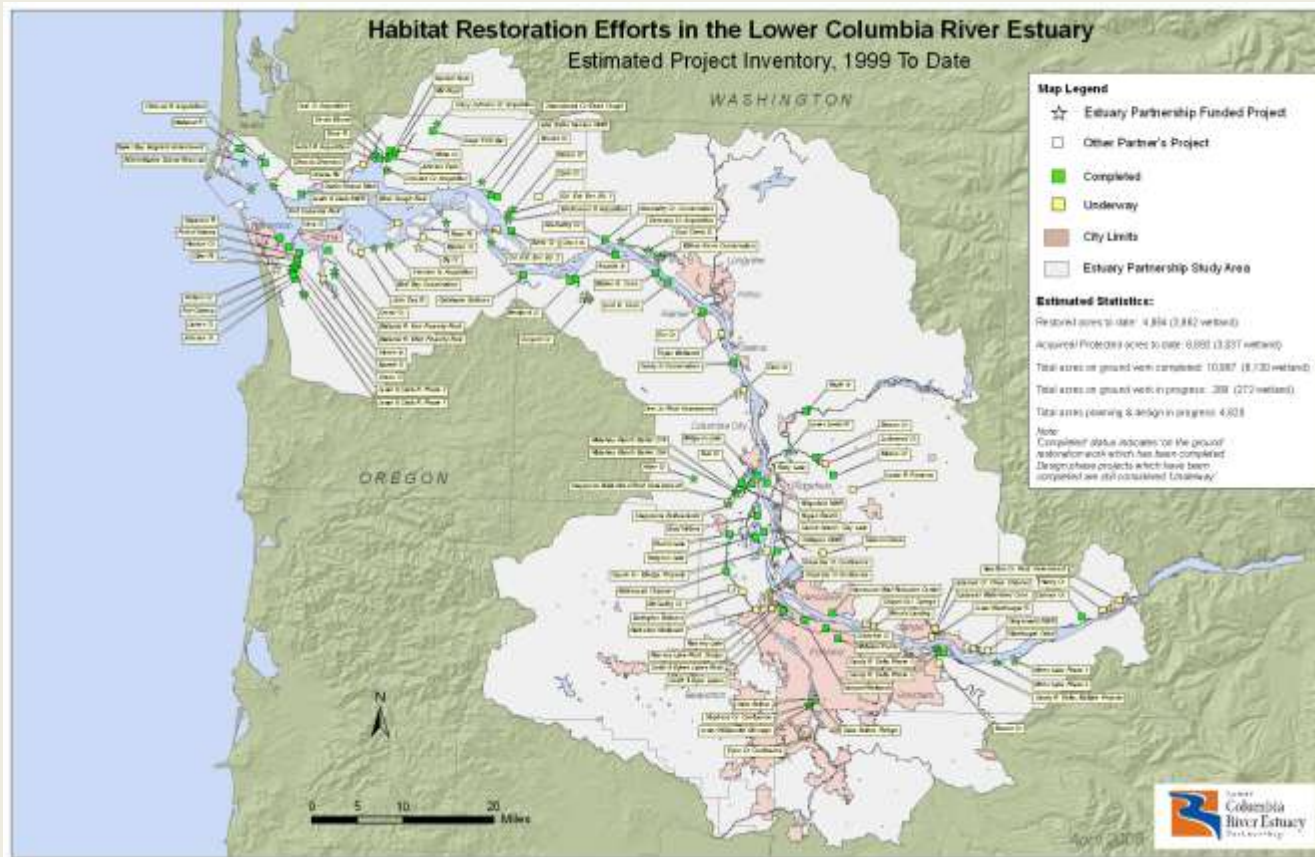




**FEEDING ECOLOGY OF JUVENILE CHINOOK SALMON
EMIGRATING THROUGH THE LOWER COLUMBIA RIVER
AND ESTUARY**

Angie Munguia
Jessica Miller
Laurie Weitkemp

BACKGROUND- SALMON RECOVERY



- Over 90% of land in the lower Columbia River and estuary has been developed
- Focus on restoring estuarine habitats
- Currently, the Lower Columbia Estuary Partnership has restored/protected over 22,000 with the goal of 25,000 wetland acres by 2025
- Breaching levees and dikes to restore wetlands
- Improving salmonid habitat (rearing, spawning, feeding)

WHICH SPECIES IS THE ESTUARY BEING RESTORED FOR?

- **Lower Columbia River estuary is an important habitat for many juvenile salmonid stocks**
- **Duration in estuary can differ by stock during emigration**
- **Most research is focused on shallow water habitat and wetland use**
- **Less is known about habitat use by threatened interior spring stocks**

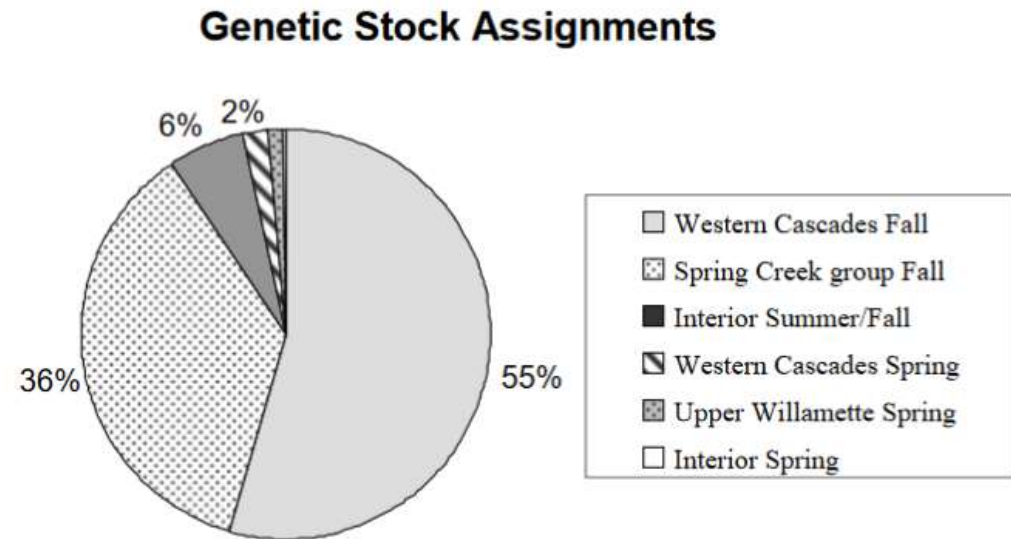
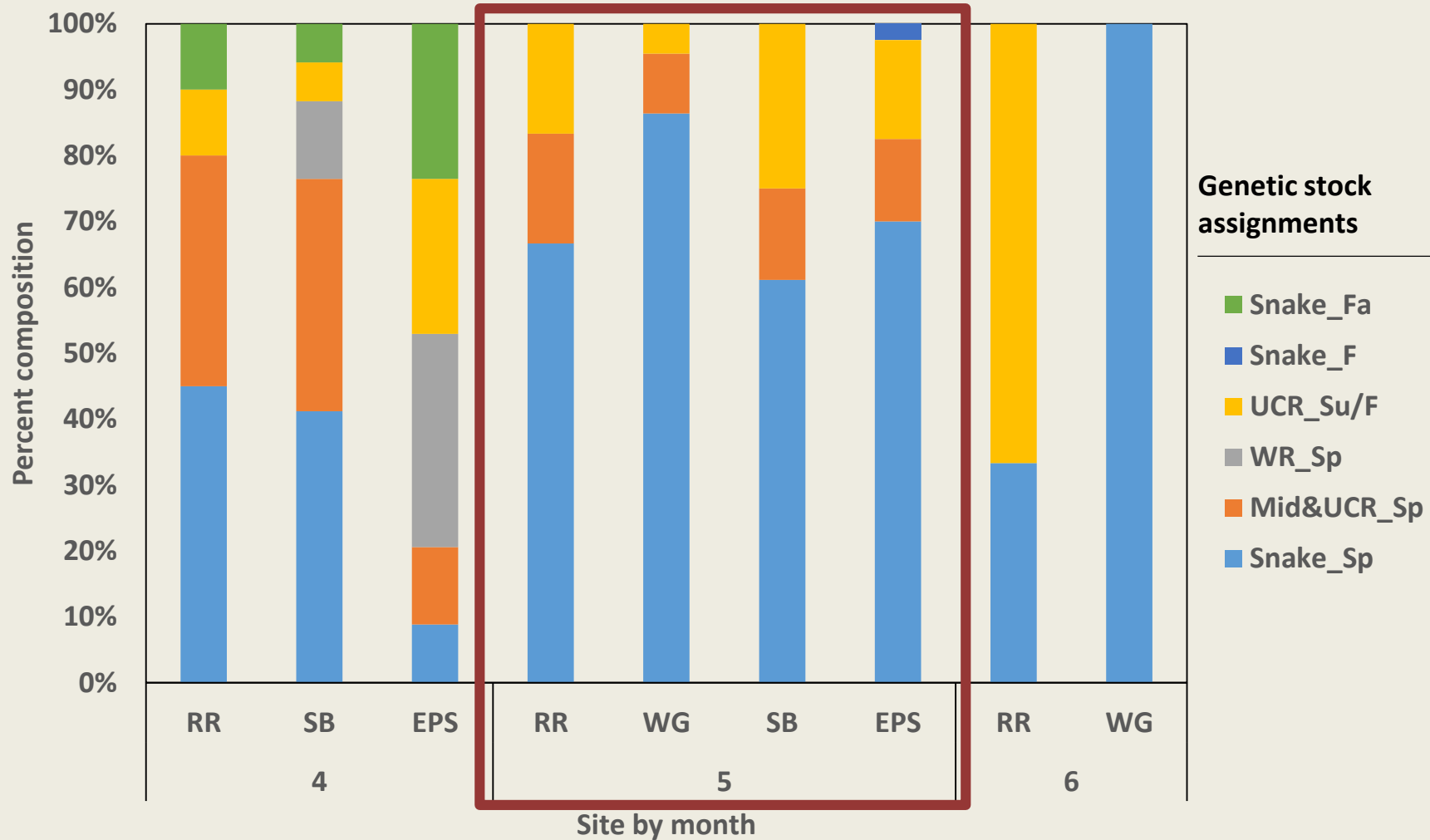


Figure 6. Estimated proportions of 6 major Chinook salmon stock groups from seven beach-seining sites in the lower Columbia River estuary, 2002-2004 (n = 1,004).

Bottom et al. 2008

MAINSTEM CATCH DATA (2016)



BIG QUESTION

- Are the estuary habitat restoration actions achieving expected biological and environmental benefits?
 - Specifically for interior stocks of Chinook salmon

Is restoration working and can we detect it?

River

Floodplain



Photo credit: Jeff Operman, UC Davis Watershed Sciences

RESEARCH OBJECTIVES

Goal: Evaluate the ecological benefits of restoration actions for yearling salmon in the lower Columbia River and estuary

OBJECTIVES:

- 1) *Characterize food habits of emigrating yearling Chinook in the LCR&E*
 - Determine diet composition and stomach fullness trends along the longitudinal continuum
- 2) *Identify if we can detect changes in food sources (stable isotope signatures) supporting emigrating yearlings*
 - *If so, can we provide any information on those changes*

METHODS: FIELD COLLECTIONS (2016-2017)

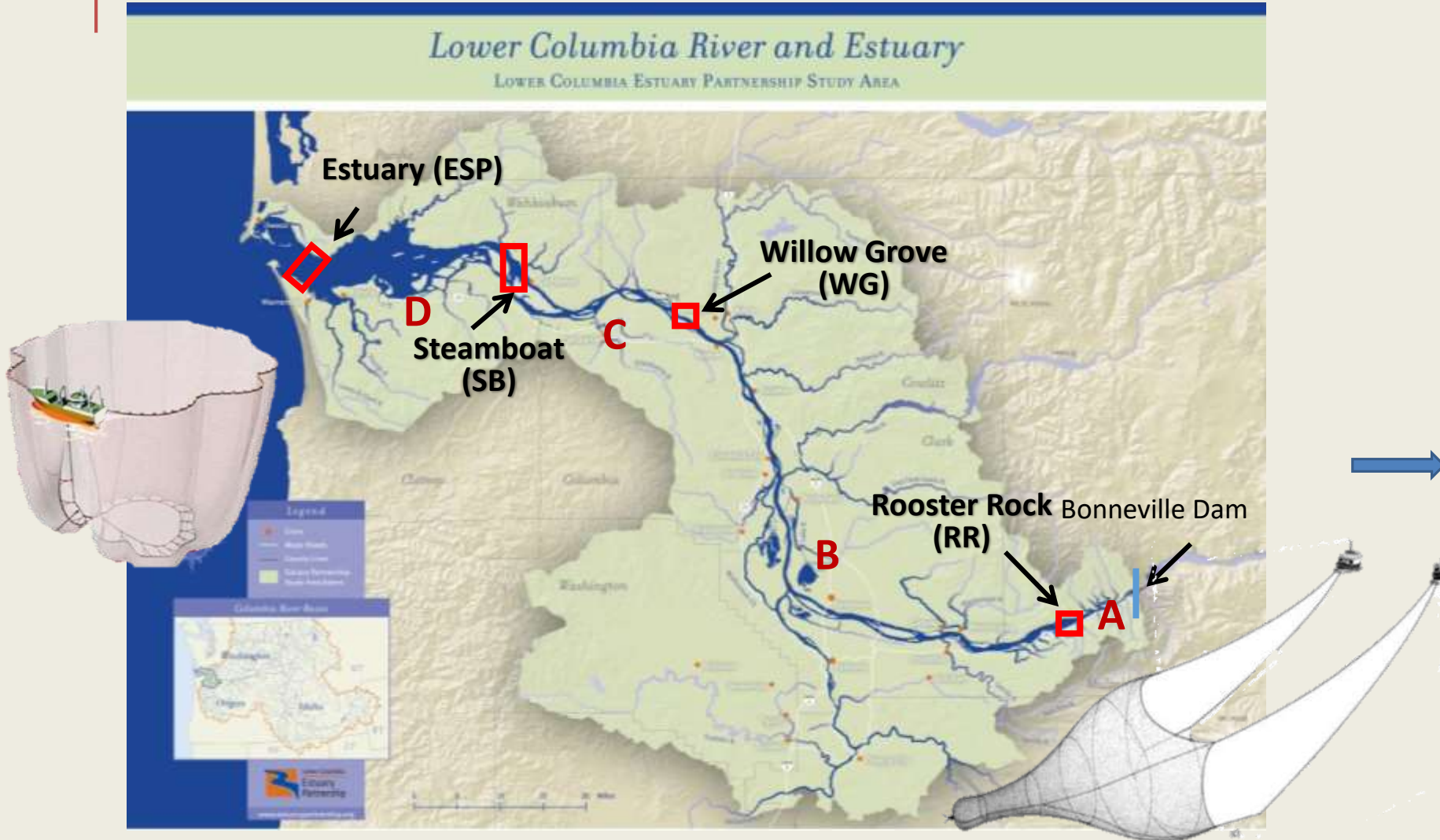
2017 only

→ McCall Hatchery Sampling

- 50 fish
- 5 tissue types:
 - Caudal fin
 - Muscle
 - Liver
 - Plasma
 - Red blood cells

→ Ocean (BPA)

- 24 fish



ANALYSES

Diet analysis (<30hrs)

- **Presence/absence of insects**

Stable isotope analysis

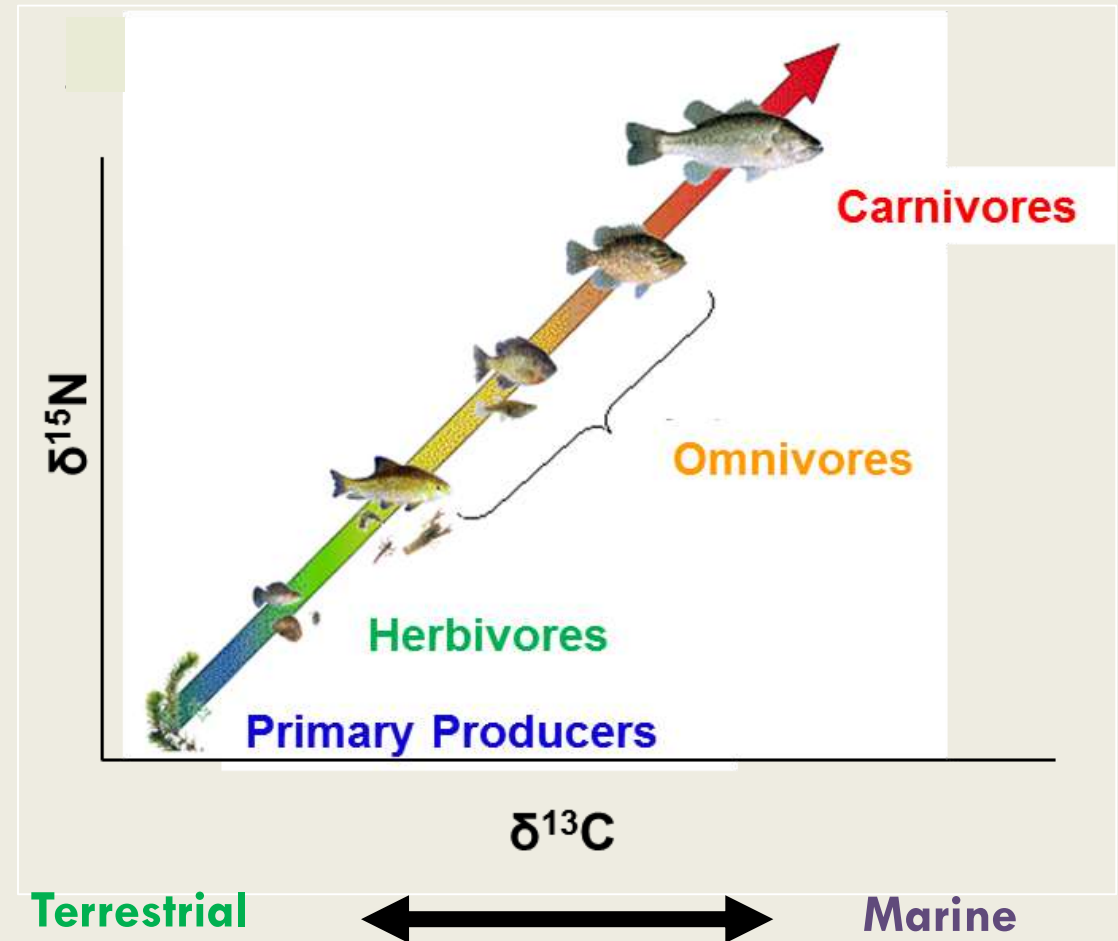
- **Tissue (Fin and Muscle) - recent food source switches**
- **Prey in stomach – stable isotope signatures of prey consumed**

For 2016

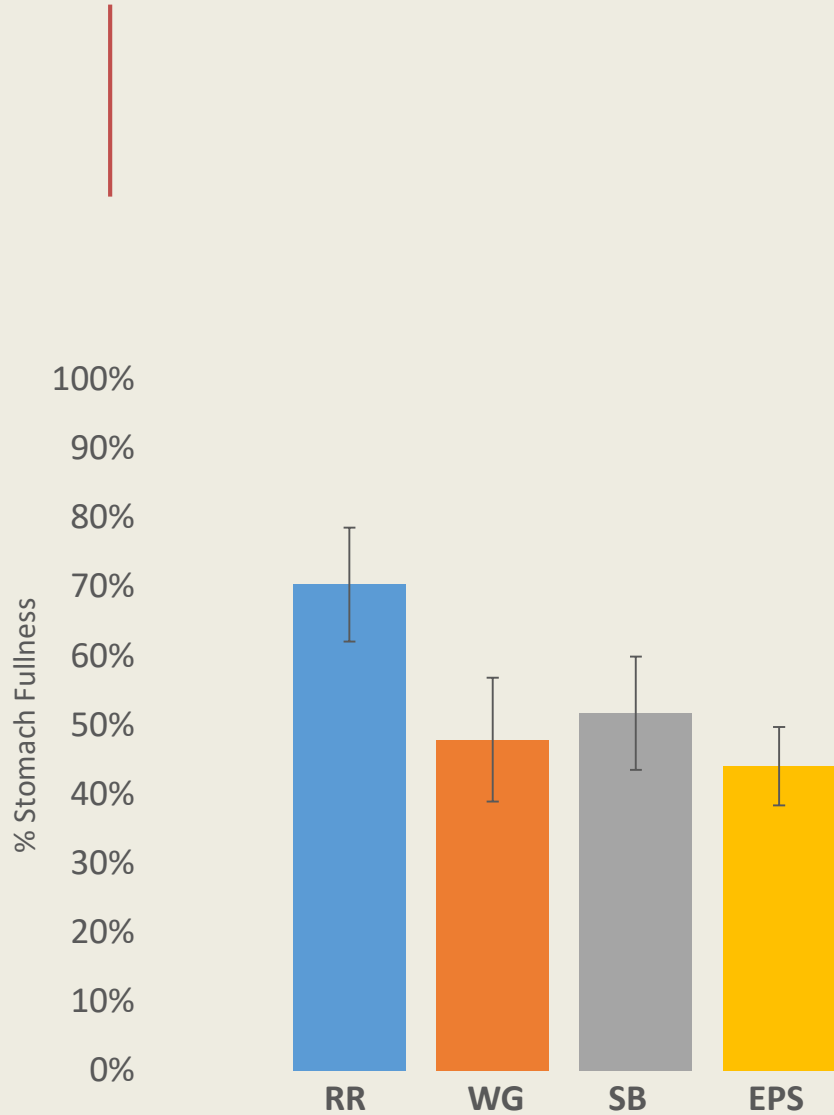


STABLE ISOTOPE ANALYSIS (SIA)

- **Powerful tool in diet and food web studies**
 - “You are what you eat!”
- **Measured as a ratio of heavy to light isotope**
- **Carbon** – Primary producers (carbon sources)
- **Nitrogen** - Trophic position
~3.5% enrichment for each trophic position



PRELIMINARY RESULTS



| | | RR | WG | SB | EPS | |
|-------------------------|---------------------------|-----------------|----|----|-----|---|
| Insecta | | | | | | |
| Coleoptera | Coleoptera | 4 | 4 | 4 | 4 | |
| | Coccinellidae | 2 | 2 | | | |
| | Dytiscid | | | 1 | | |
| | Elmidae | | 1 | | | |
| | Staphylinidae | 1 | | | | |
| | Diptera | Diptera | 4 | 4 | 4 | 4 |
| | | Cecidomyiidae | | | 1 | |
| | | Ceratopogonidae | | | 2 | 2 |
| | | Chironomidae | 4 | 4 | 4 | 4 |
| | Simuliidae | | 1 | | | |
| | Diptera-Brachycera | Muscidae | | | 1 | |
| | | Ephemeroptera | 3 | 3 | 3 | |
| | Hemiptera | Hemipteran | 2 | | 2 | |
| | | Aphididae | 3 | | 3 | 3 |
| | | Cicadellidae | 1 | | | |
| | | Psyllidae | | | 1 | |
| | | Tingidae | | 1 | | |
| | Hymenoptera | Hymenoptera | 4 | 4 | 4 | 4 |
| | | Formicidae | | 1 | | |
| Odonata | Anisoptera | | 1 | | | |
| | Coenagrionidae | | 1 | | | |
| Plecoptera | Zygoptera | 2 | | 2 | | |
| | Plecoptera | 1 | | | | |
| Trichoptera | Trichoptera case | 2 | | | 2 | |
| | Insect | 4 | 4 | 4 | 4 | |
| Other arthropoda | | | | | | |
| | Acari | | | | 1 | |
| | Araneae | 3 | 3 | 3 | | |
| Crustacea | | | | | | |
| Amphipod | Amphipod | 4 | 4 | 4 | 4 | |
| | Americorophium | 4 | 4 | 4 | 4 | |
| Corophiidae | <i>A. salmonis</i> | | 2 | 2 | | |
| | <i>A. spinicornis</i> | | 2 | 2 | | |
| | Gammaridae | 4 | 4 | 4 | 4 | |
| | Orange gammarus | 3 | 3 | 3 | | |
| | <i>G. lacustris</i> | 3 | 3 | 3 | | |
| Cladocera | Daphnia sp | | | 2 | 2 | |
| | Crustacean parts | 3 | 3 | | 3 | |
| Other crustacea | Cirripedia | 2 | | | 2 | |
| | Mysidae | 1 | | | | |
| Other | | | | | | |
| | Bivalve | 3 | | 3 | 3 | |
| | Fish scale | 4 | 4 | 4 | 4 | |
| | Nematoda | | 3 | 3 | 3 | |
| | Rocks/seeds/plants | 4 | 4 | 4 | 4 | |
| | Plastic | | 2 | 2 | | |
| | Unidentified matter | 4 | 4 | 4 | 4 | |

PRELIMINARY RESULTS



Hymenopteran



Americorophium

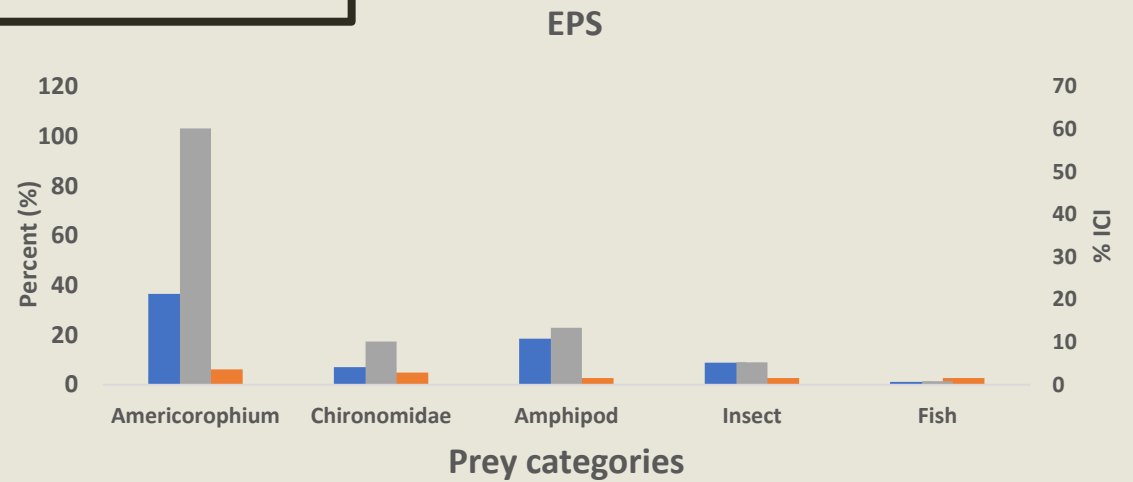
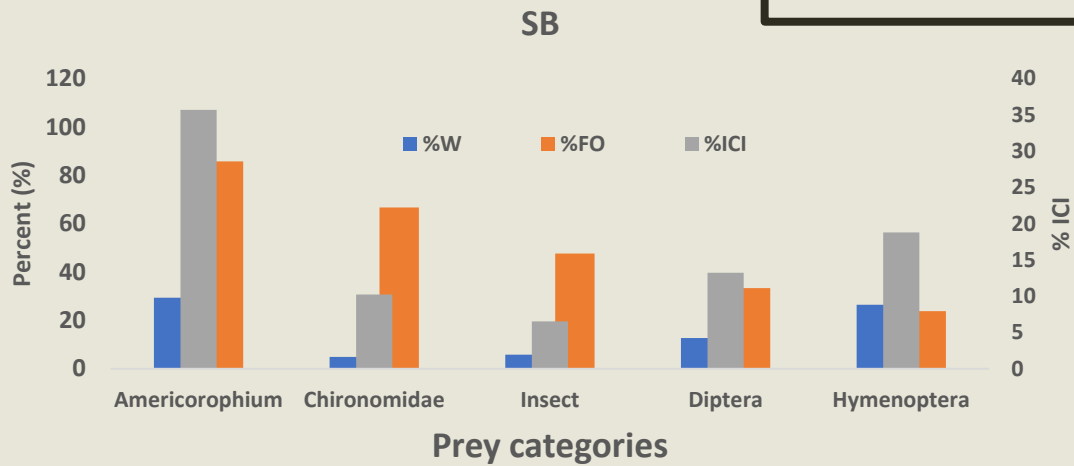
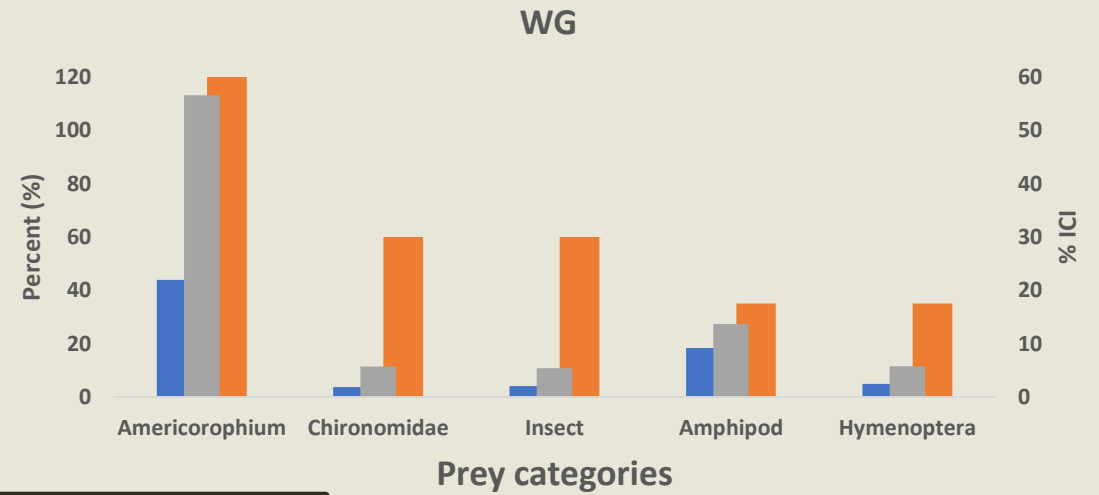
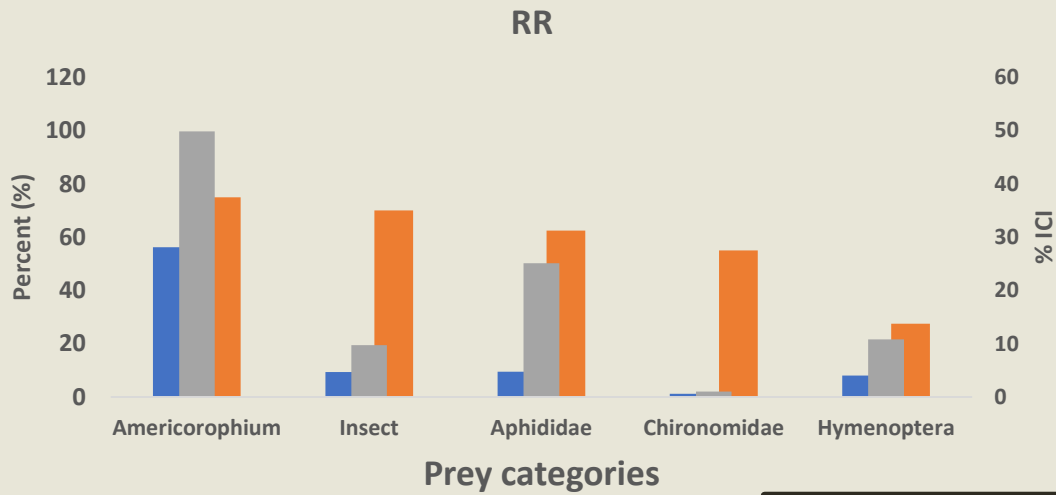
| Insecta | | RR | WG | SB | EPS | |
|---------------------------|---------------------|-----------------|----|----|-----|---|
| Coleoptera | Coleoptera | 4 | 4 | 4 | 4 | |
| | Coccinellidae | 2 | 2 | | | |
| | Dytiscid | | | 1 | | |
| | Elmidae | | 1 | | | |
| | Staphylinidae | 1 | | | | |
| | Diptera | Diptera | 4 | 4 | 4 | 4 |
| | | Cecidomyiidae | | | 1 | |
| | | Ceratopogonidae | | | 2 | 2 |
| | | Chironomidae | 4 | 4 | 4 | 4 |
| | | Simuliidae | | 1 | | |
| Diptera-Brachycera | Muscidae | | | 1 | | |
| | Ephemeroptera | 3 | 3 | 3 | | |
| Hemiptera | Hemipteran | 2 | | 2 | | |
| | Aphididae | 3 | | 3 | 3 | |
| | Cicadellidae | 1 | | | | |
| | Psyllidae | | | 1 | | |
| | Tingidae | | 1 | | | |
| Hymenoptera | Hymenoptera | 4 | 4 | 4 | 4 | |
| | Formicidae | | 1 | | | |
| Odonata | Anisoptera | | 1 | | | |
| | Coenagrionidae | | 1 | | | |
| | Zygoptera | 2 | | 2 | | |
| Plecoptera | Plecoptera | 1 | | | | |
| Trichoptera | Trichoptera case | 2 | | | 2 | |
| Insect | Insect | 4 | 4 | 4 | 4 | |
| Other arthropoda | | | | | | |
| | Acari | | | | 1 | |
| | Araneae | 3 | 3 | 3 | | |
| Crustacea | | | | | | |
| Amphipod | Amphipod | 4 | 4 | 4 | 4 | |
| | Corophiidae | 4 | 4 | 4 | 4 | |
| Gammaridae | Gammaridae | 4 | 4 | 4 | 4 | |
| | Orange gammarus | 3 | 3 | 3 | | |
| | <i>G. lacustris</i> | 3 | 3 | 3 | | |
| Cladocera | Daphnia sp | | | 2 | 2 | |
| Other crustacea | Crustacean parts | 3 | 3 | | 3 | |
| | Cirripedia | 2 | | | 2 | |
| | Mysidae | 1 | | | | |
| Other | | | | | | |
| | Bivalve | 3 | | 3 | 3 | |
| | Fish scale | 4 | 4 | 4 | 4 | |
| | Nematoda | | 3 | 3 | 3 | |
| | Rocks/seeds/plants | 4 | 4 | 4 | 4 | |
| | Plastic | | 2 | 2 | | |
| | Unidentified matter | 4 | 4 | 4 | 4 | |

PRELIMINARY RESULTS



Odonata: Coenagrionidae

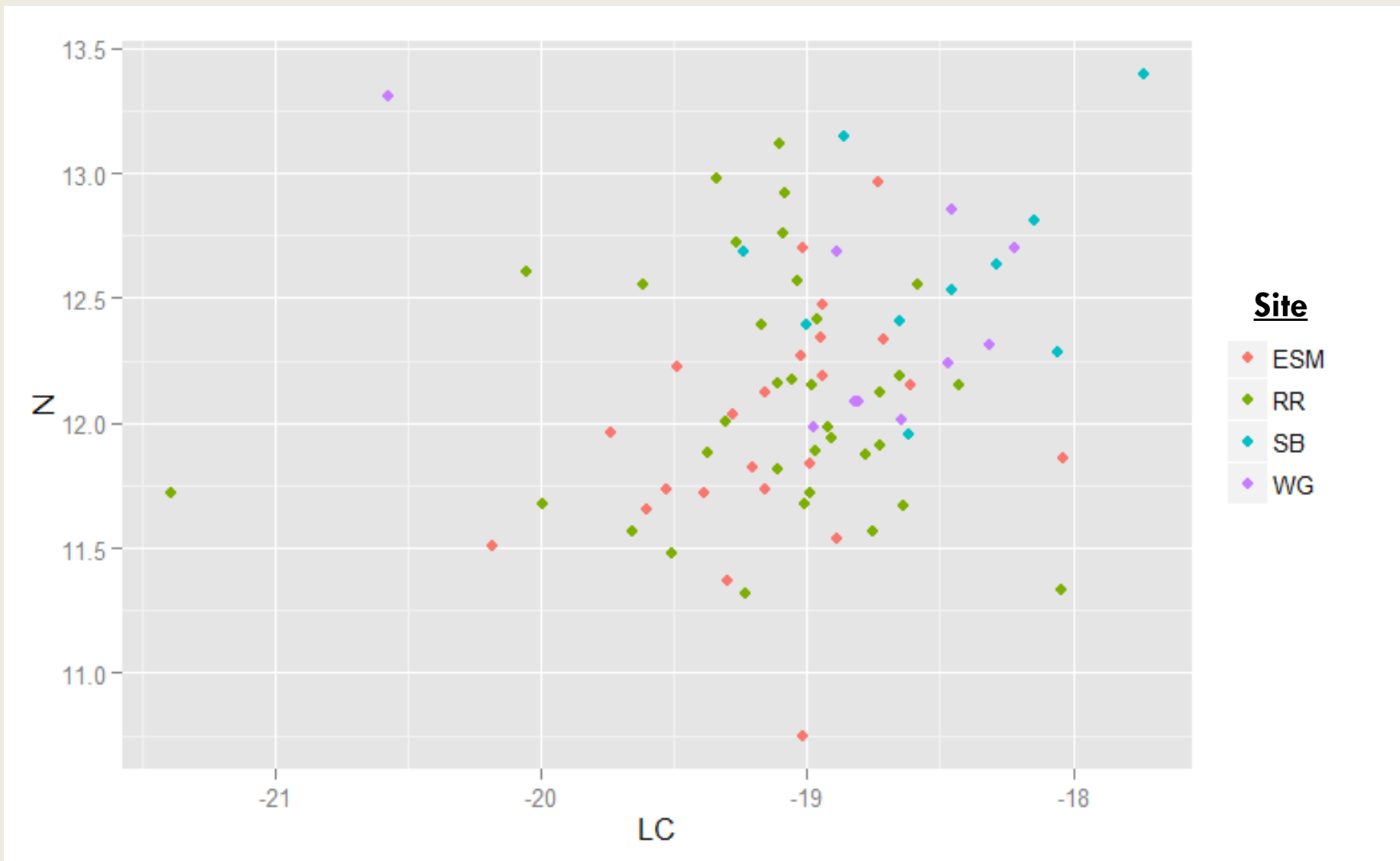
| Insecta | | RR | WG | SB | EPS | |
|------------------------|---------------------------|-----------------|----|----|-----|---|
| Coleoptera | Coleoptera | 4 | 4 | 4 | 4 | |
| | Coccinellidae | 2 | 2 | | | |
| | Dytiscid | | | 1 | | |
| | Elmidae | | 1 | | | |
| | Staphylinidae | 1 | | | | |
| | Diptera | Diptera | 4 | 4 | 4 | 4 |
| | | Cecidomyiidae | | | 1 | |
| | | Ceratopogonidae | | | 2 | 2 |
| | | Chironomidae | 4 | 4 | 4 | 4 |
| | | Simuliidae | | 1 | | |
| | Diptera-Brachycera | Muscidae | | | 1 | |
| | Ephemeroptera | Ephemeroptera | 3 | 3 | 3 | |
| | Hemiptera | Hemipteran | 2 | | 2 | |
| Aphididae | | 3 | | 3 | 3 | |
| Cicadellidae | | 1 | | | | |
| | Psyllidae | | | 1 | | |
| | Tingidae | | 1 | | | |
| Hymenoptera | Hymenoptera | 4 | 4 | 4 | 4 | |
| | Formicidae | | 1 | | | |
| Odonata | Anisoptera | | 1 | | | |
| | Coenagrionidae | | 1 | | | |
| | Zygoptera | 2 | | 2 | | |
| Plecoptera | Plecoptera | 1 | | | | |
| Trichoptera | Trichoptera case | 2 | | | 2 | |
| Insect | Insect | 4 | 4 | 4 | 4 | |
| Other arthropoda | | | | | | |
| | Acari | | | | 1 | |
| | Araneae | 3 | 3 | 3 | | |
| Crustacea | | | | | | |
| Amphipod | Amphipod | 4 | 4 | 4 | 4 | |
| Corophiidae | Americorophium | 4 | 4 | 4 | 4 | |
| | <i>A. salmonis</i> | | 2 | 2 | | |
| | <i>A. spinicornis</i> | | 2 | 2 | | |
| Gammaridae | Gammaridae | 4 | 4 | 4 | 4 | |
| | Orange gammarus | 3 | 3 | 3 | | |
| | <i>G. lacustris</i> | 3 | 3 | 3 | | |
| | Daphnia sp | | | 2 | 2 | |
| Cladocera | Daphnia sp | | | 2 | 2 | |
| Other crustacea | Crustacean parts | 3 | 3 | | 3 | |
| | Cirripedia | 2 | | | 2 | |
| | Mysidae | 1 | | | | |
| Other | | | | | | |
| | Bivalve | 3 | | 3 | 3 | |
| | Fish scale | 4 | 4 | 4 | 4 | |
| | Nematoda | | 3 | 3 | 3 | |
| | Rocks/seeds/plants | 4 | 4 | 4 | 4 | |
| | Plastic | | 2 | 2 | | |
| | Unidentified matter | 4 | 4 | 4 | 4 | |



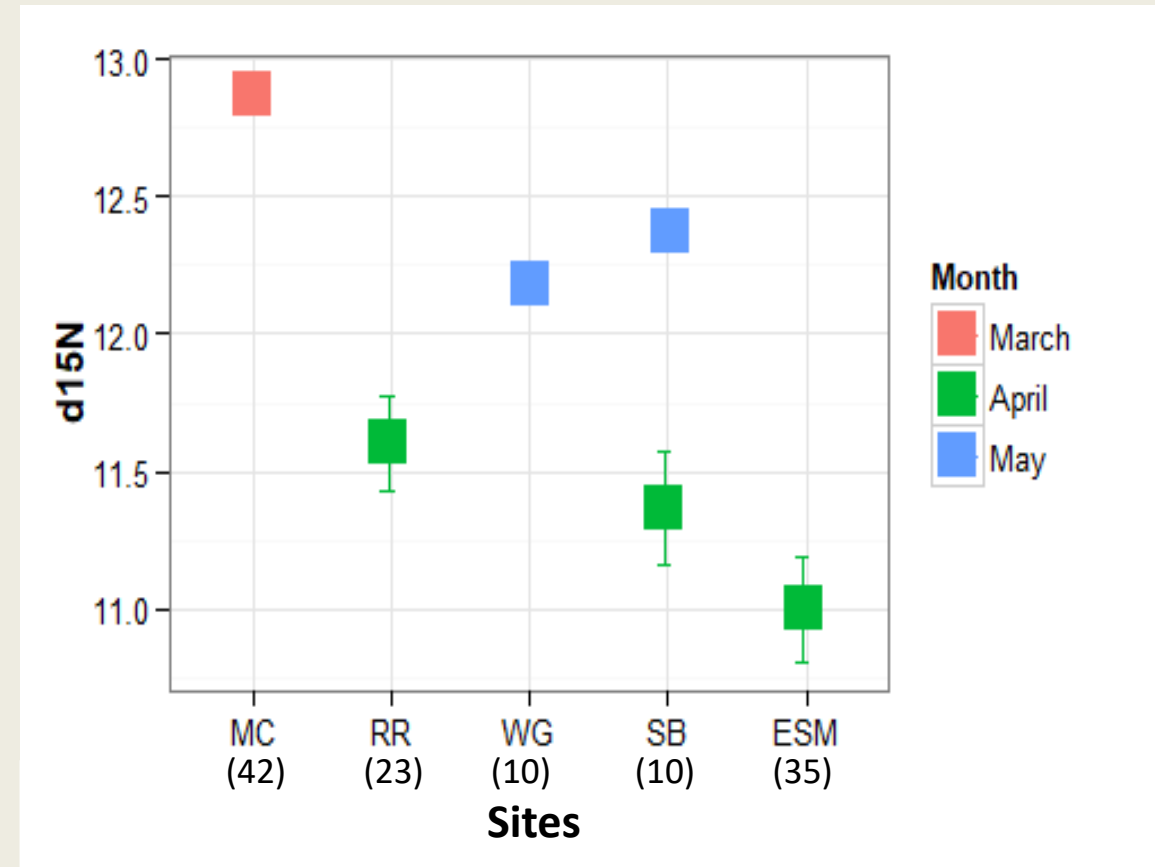
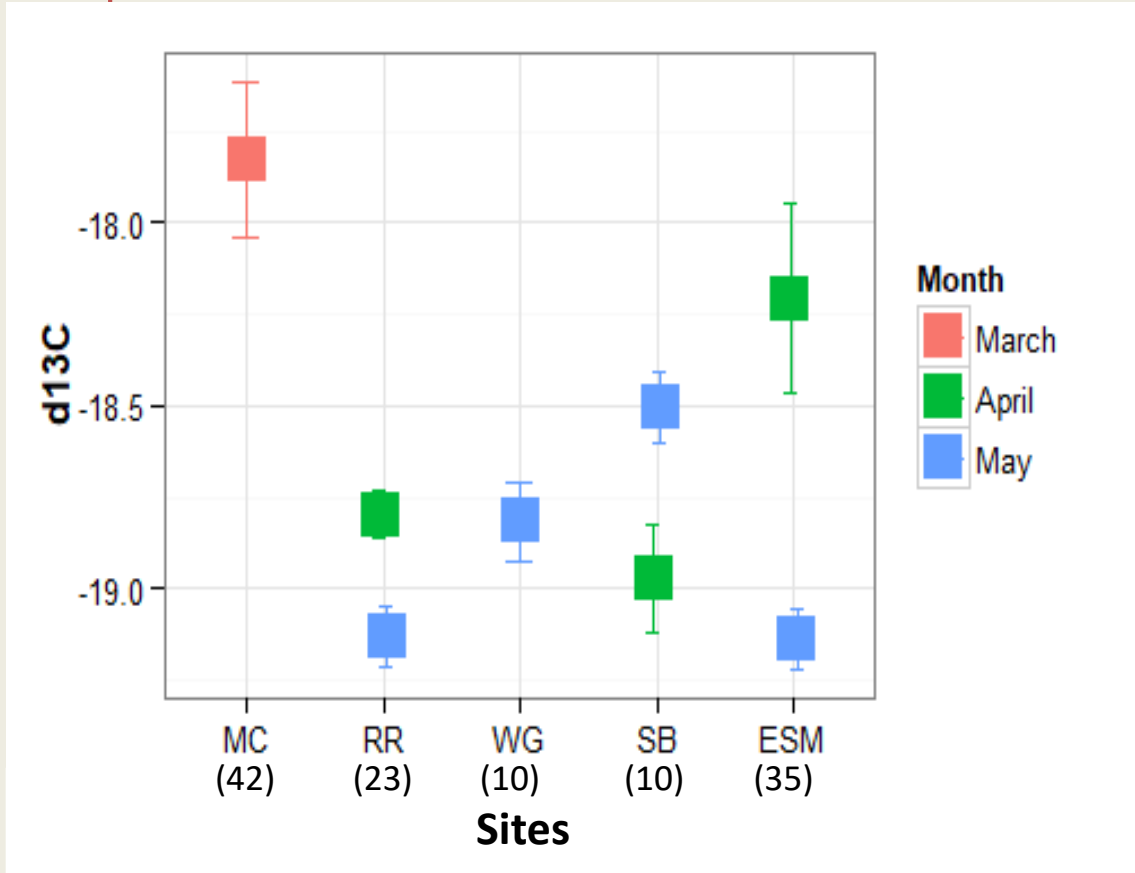
IMPORTANT PREY BY SITE



FIN SAMPLES (MAY 2016)



FIN STABLE ISOTOPE (MAY 2016)



Dam



Ocean

SIGNIFICANCE: MAINSTEM SPRING CHINOOK SIA

Carbon (d13C)

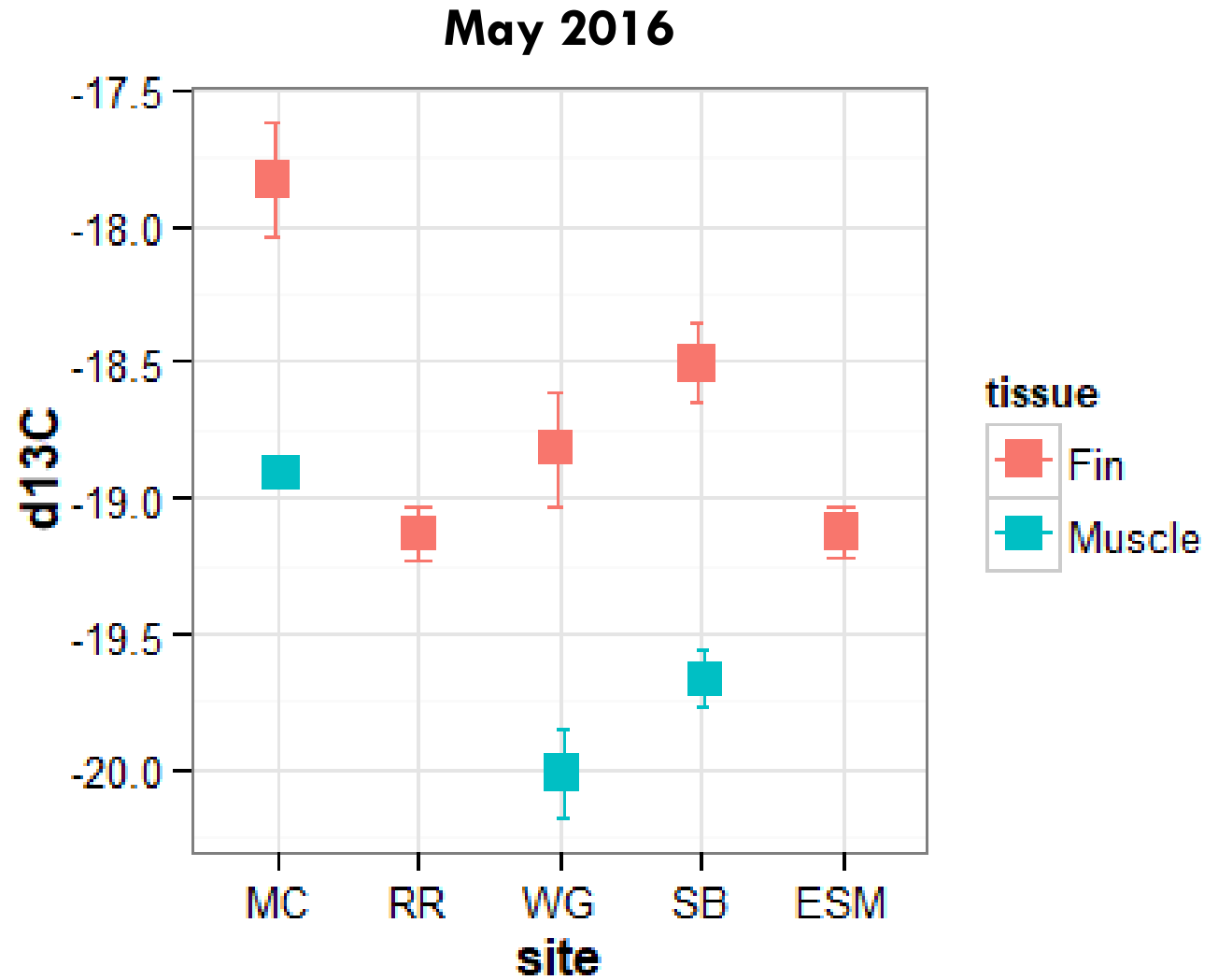
- SB and RR ($p < 0.05$)
- SB and ESP ($p < 0.05$)
- McCall is significantly different from all sites except SB

Nitrogen (d15N)

- SB and RR ($p < 0.05$)
- SB and ESP ($p < 0.05$)
- WG and ESP (< 0.05)
- McCall is significantly from ESP and RR

STABLE ISOTOPE SIGNATURE BY TISSUE TYPE

- Mean shift in muscle and fin stable isotope signatures ~1 unit
- Smaller sample size for muscle
- No samples collected from RR and ESM



WHAT IS NEXT?

- Using McCall samples and possibly prey and primary source data from monitoring sites in the LCR&E to aid in interpretation of field samples
- Stable isotope analysis on most common prey items from yearling stomach contents
- Determine if there is variability within the caudal fin tissue
 - Split fin into thirds
- 2017 AEMR samples + BPA Ocean samples
- Compare yearling Chinook data to steelhead and sub-yearling fin data (2017)



ACKNOWLEDGEMENTS



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Jessica Miller

Field Crew and Boat Operators

Jake Biron, Wayne Haines, Lance Renoux, Brian Kelly, Kaya Johnson

McCall Collections

Beckman Lab, Peik Anderesen

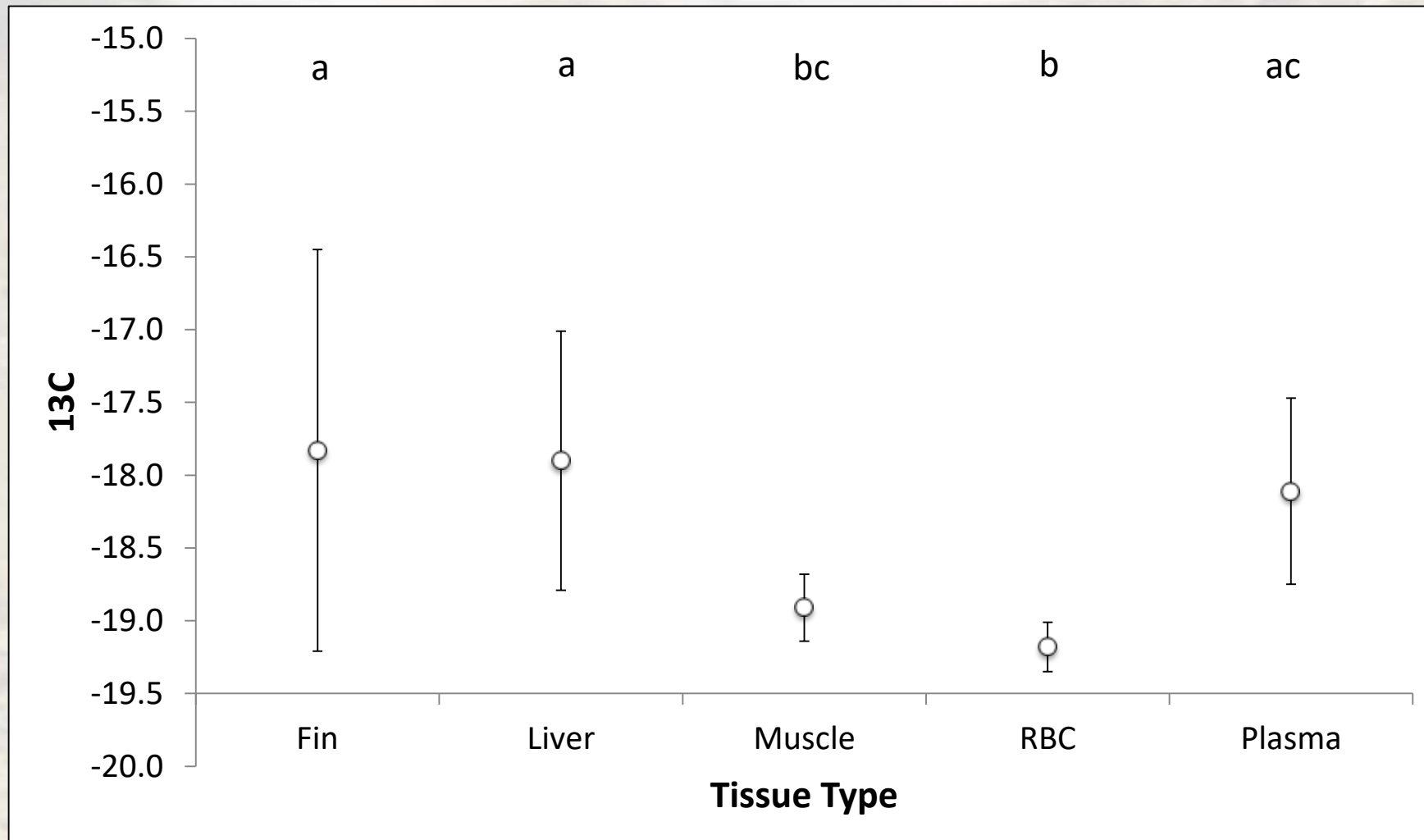
Funding

NOAA LMRCSC and EPP, Mamie Markham Research Award

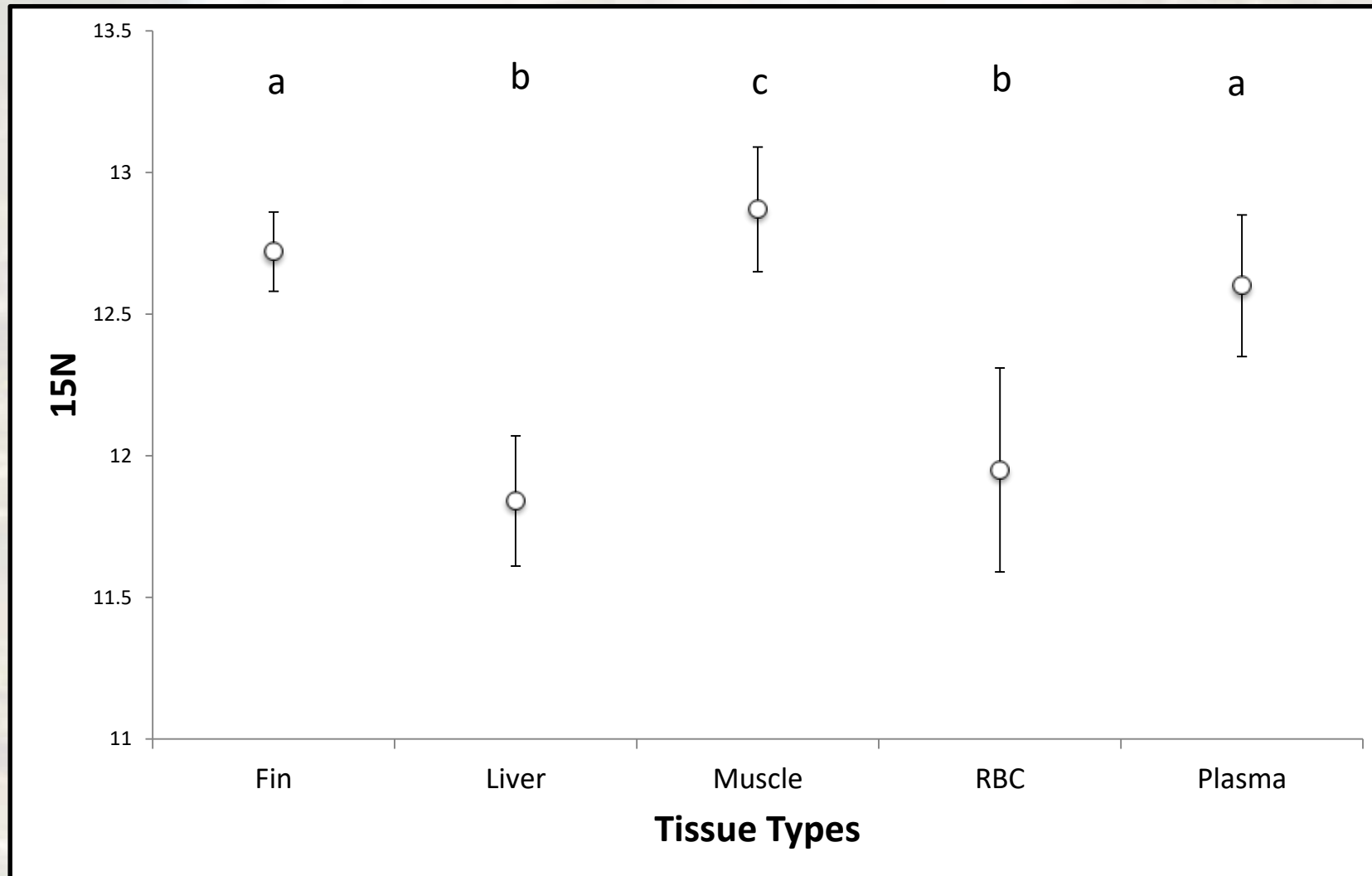
Interns

Sawyer Finley, Elle Bowman

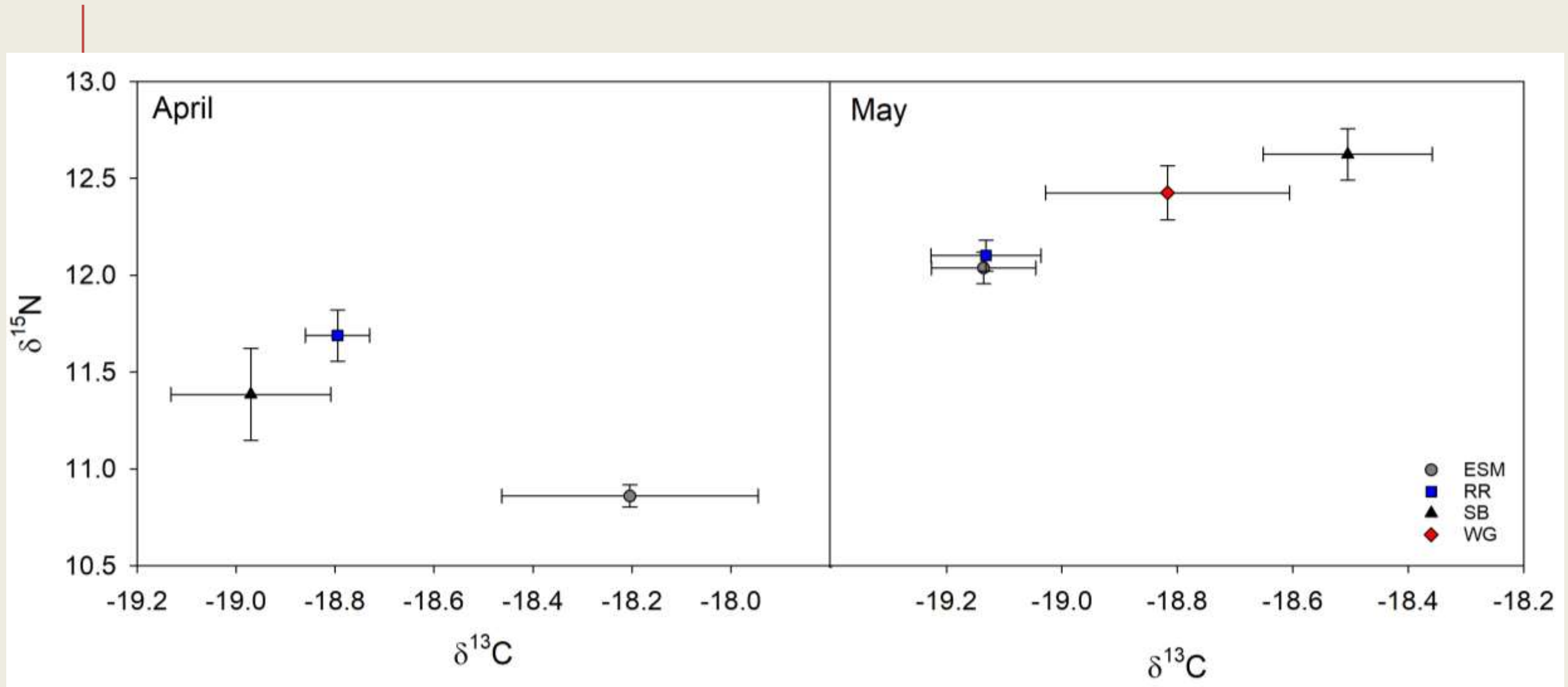
Lipid corrected $\delta^{13}\text{C}$ values differed significantly between tissue types



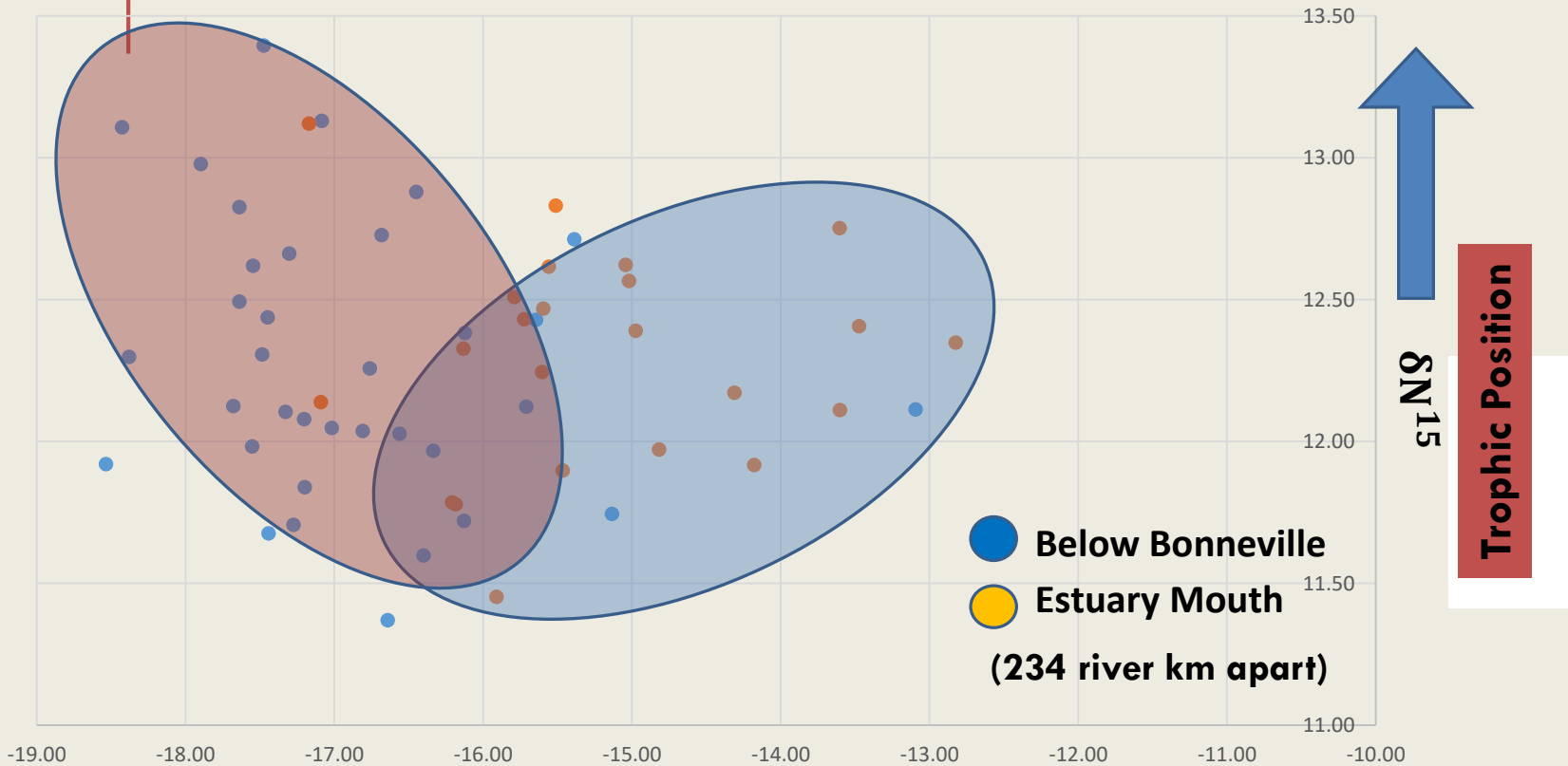
$\delta^{15}\text{N}$ rates were different between most tissue types



Fin Tissue



PRELIMINARY: SIA FIN TISSUE



- Snake Spring Chinook salmon collected in May
- Variable migration rates and food sources seen in overlap
- Moving away from hatchery signal (decrease in N)
- Diet switches occurring during emigration
- Middle sites and April samples yet to be analyzed



Dam



δC^{13}



Ocean

| | P-value |
|------|---------|
| C:N | < 0.01 |
| dN15 | 0.88 |
| dC13 | <0.01 |

CHALLENGES AND LIMITATIONS

- Will fin tissue have a fast enough turnover rate that I can identify shifts in food sources during their emigration through the LCR&E?
- How well am I going to be able to translate my stable isotope signatures into actual sources?

METHODS: DIET ANALYSIS



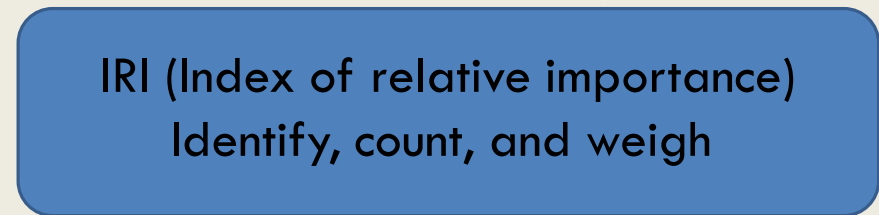
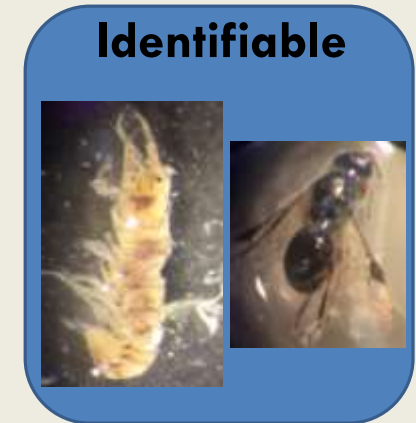
Analysis:

Diet composition

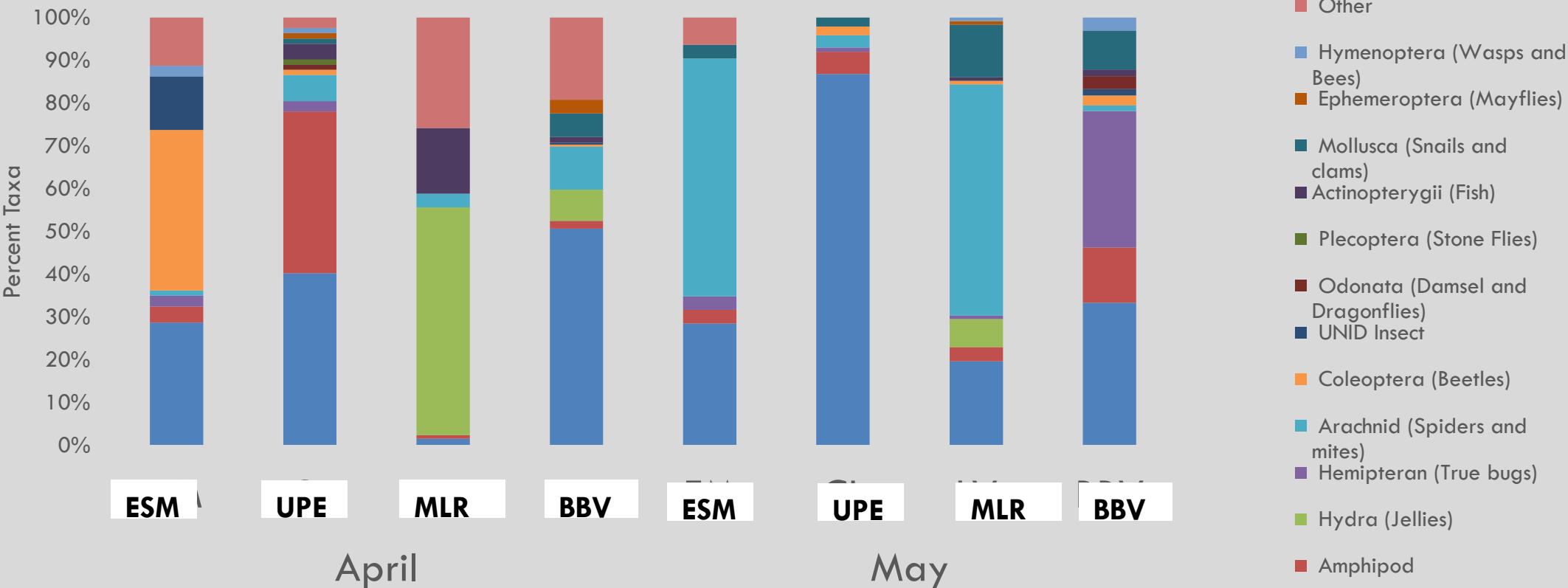
- % similarity indices (between sites)
- NMDS (Environmental variables, site, year, month)

Stomach fullness

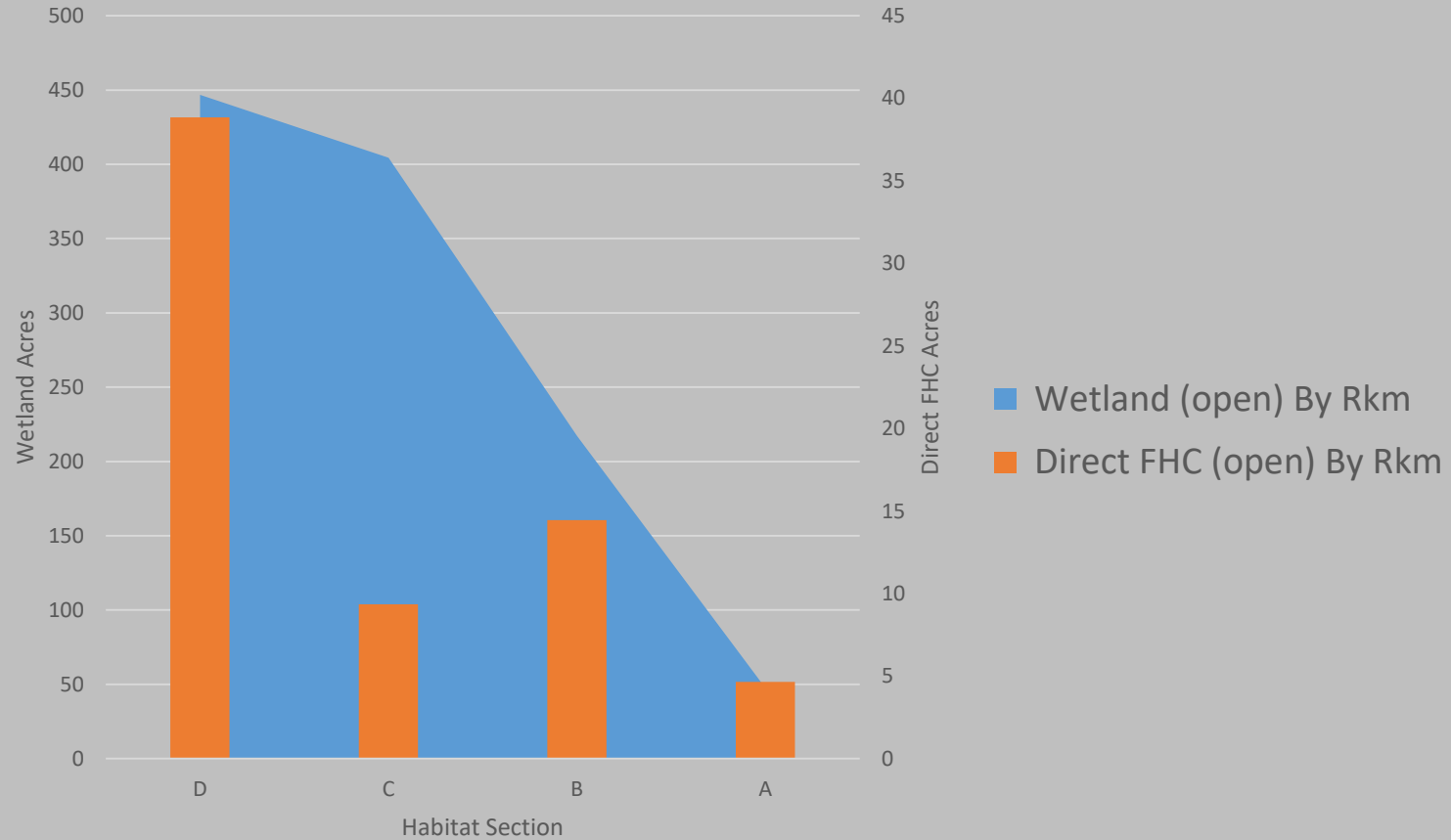
- Two-way ANOVA (between sites, months)



NEUSTON

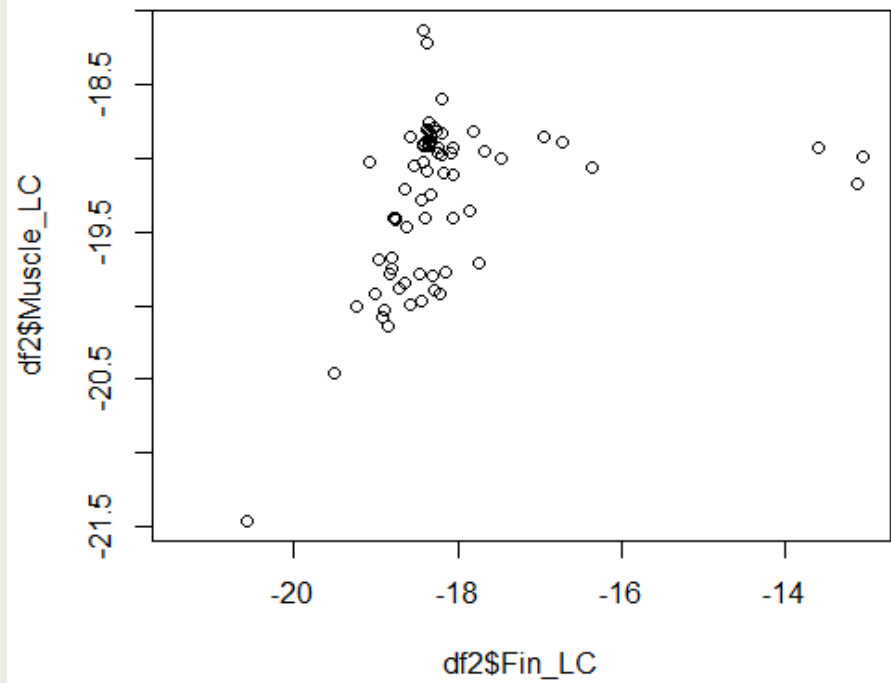


WETLAND AND FISH HABITAT BY RKM

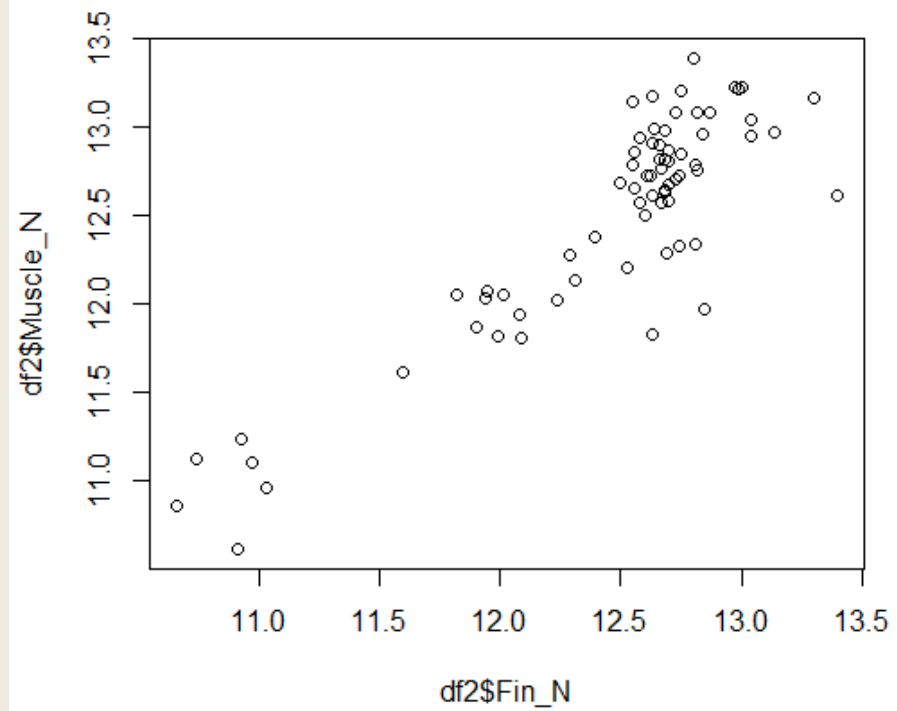


NEED TO ID





$r = 0.7732904$



$r = 0.9220092$